

Comparison of *bombyx mori* L. Fibroin coatings on the mechanical properties of collagen membranes

Helmi Fathurrahman*, **, Siti Sunarintyas***, Sri Budi Barunawati****, Nuryono*****

* Doctorate Student Faculty of Dentistry Gadjah Mada University Yogyakarta

** Departement of Prosthodontics, Faculty of Dentistry, Sultan Agung Islamic University, Semarang

***Department of Dental Materials, Faculty of dentistry, Gadjah Mada University, Yogyakarta

****Department of Prosthodontics, Faculty of dentistry, GadjahMada University, Yogyakarta

**** Department of Chemistry, Faculty of Mathematics and Natural Sciences, Gadjah Mada University, Yogyakarta

Correspondence: helmi_f@unissula.ac.id

Received 02 April 2023; 1st revision 12 May 2023; 2nd revision 12 July 2023; Accepted 26 July 2023;

Published online 31 July 2023

Keywords:

Collagen membrane;
Bombyx mori L. fibroin;
guided bone regeneration.

ABSTRACT

Background: Most of dental implants are accompanied by guided bone regeneration (GBR) procedures. Collagen membrane which is the most widely used type of membrane in the GBR procedure has the disadvantage of not having antimicrobial properties, old bone formation, easy to tear, and soft so that it does not maintain the dimensions of the augmentation area. *Bombyx mori* L. fibroin has good mechanical properties and has the potential to be used as a pharmaceutical material that is biocompatible, antimicrobial, and stimulates bone growth. The aim of this study was to analyze the effect of *Bombyx mori* L. fibroin coating on the mechanical properties of collagen membranes.

Method: The samples consisted of the control group (group A) and the treatment group, the collagen membrane was coated using the *Bombyx mori* L. dip-coating fibroin technique, namely group B (1 time coating process), group C (2 times coating process) and group D (3 times the coating process). Statistical analysis used one-way ANOVA ($p < 0.05$).

Result: The results showed that the average tensile strength of the negative control group was 2.62 MPa, the 1-coated group was 4.51 MPa, the 2-coated group was 5.45 MPa, and the 3-coated group was 1.41. The significance value of ANOVA was 0.000, the post hoc test results showed that there were significant differences in all testgroups with a significance level of 0.005

Conclusion: It is concluded that coating *Bombyx mori* L. fibroin on collagen membranes increases the physicommechanical properties of tensile strength.

Copyright ©2022 National Research and Innovation Agency. This is an open access article under the CC BY-SA license (<https://creativecommons.org/licenses/by-sa/4.0/>).

doi: <http://dx.doi.org/10.30659/odj.10.1.140-146>

2460-4119 / 2354-5992 ©2022 National Research and Innovation Agency

This is an open access article under the CC BY-SA license (<https://creativecommons.org/licenses/by-sa/4.0/>)

Odonto : Dental Journal accredited as Sinta 2 Journal (<https://sinta.kemdikbud.go.id/journals/profile/3200>)

How to Cite: Fathurrahman *et al.* Comparison of *bombyx mori* L. Fibroin coatings on the mechanical properties of collagen membranes. Odonto: Dental Journal, v.10, n.1, p. 140-146, July 2023

INTRODUCTION

The prevalence of tooth loss based on the results of the national health research, is 51.4%. (1). Replacement of missing teeth is using a denture that aims to restore the function of mastication, phonation, providing facial muscle support in supporting appearance and maintaining the remaining teeth (2). Based on how to wear a denture consists of a removable denture and a fixed denture. According to Zitzmann *et al*, (3) the number of users of fixed dentures ranges from 31%-45% of all denture users. The number of dental implant users has increased sharply, from 0.7% in 2000 to 5.7% in 2016 (4). The advantages of using dental implants are comfortable use, long wearability, excellent aesthetics, biocompatible with oral cavity tissues (5).

Research shows that more than 50% of dental implant installations require a process of alveolaris growth procedure. One of the procedures for growing the process of alveolaris is guided bone regeneration (GBR). (6,7). GBR is a bone regeneration procedure using barrier membrane with *bone graft* material. The application of GBR can increase the success of dental implant treatment up to 93.8% (8).

Based on the ability of absorption by the body, the barrier membrane used in the GBR procedure consists of 2 types, namely non-resorbable and resorbable membrane. Non-resorbable membrane is a membrane material that cannot be absorbed by the body. Resorbable membrane is a membrane material that can be absorbed by the body. Examples of this material are polycaprolactone (PCL), chitosan membrane, poly(D,L-lactic acid) (PDLLA) and collagen membrane (7). The most widely used barrier membrane material is collagen membrane (9). The disadvantages of

this material are that it does not have antimicrobial properties, long bone formation, easy to tear, and soft so that it does not maintain the dimensions of the augmentation area (10).

Bombyx mori L. is a widely cultivated species of silkworm. This species has great potential to be utilized in the health field, including as a medicine for bronchial asthma, diabetes, hepatitis, hypertension, antimicrobial, antihistamine, and bone growth spur (11,12). *Bombyx mori* cocoons have macromolecular proteins in the form of fibroin that have biocompatible, osteoinduction, anti-microbial, anti-inflammatory characteristics (13-15). Fibroin has good mechanical properties and accelerates the process of bone formation (16). It also has the ability to increase the tensile strength of medical devices in orthopedic procedures (17).

Fibroin derived from silkworms has the potential to be used as a pharmaceutical material applied by *coating* (18). Silk fibroin can be formed into thin nanoscale coatings. The thickness of this fibroin layer reaches 10 nanometers (19). Based on the description above, the author intends to conduct research on the comparison of *Bombyx mori L.* fibroin coating on the mechanical properties of collagen membranes.

MATERIALS AND METHODS

The research is laboratory experiment. The sample consisted of control group (group A) and treatment group of collagen membrane coated using *dip-coating* process of *Bombyx mori L.* fibroin, namely group B (1 time *coating* process), group C (2 times *coating* process) and group D (3 times *coating* process).

Bombyx mori L. fibroin solution with a concentration of 25 mg/mL was made by mixing

50 mg/mL fibroin 20 mL diluted using distilled water until the volume reached 40 mL. A 20 mL beaker glass was filled with fibroin *Bombyx mori L.* 25 mg/mL concentration solution as much as 13 mL. Collagen membrane measuring 2x1.5 cm was placed inside the closed beaker glass containing fibroin solution. Collagen membranes were arranged in such a way that all collagen membrane surfaces were submerged. The glass beaker that had been covered with aluminum foil was inserted in a sonicator that had been filled with distilled water until half the height of the cup was submerged. The sonicator was turned on at a frequency of 20-60 Hz, the soaking process lasted for 12 minutes. The collagen membrane was then removed and dried at room

temperature for 24 hours. The coating process was repeated for groups C, and D.

Tensile strength test was conducted using a universal testing machine. Collagen membrane samples measuring 2 cm long, 1.5 cm wide, 0.1 cm thick that had been coated with *Bombyx mori L.* fibroin solution were placed on the longitudinal side between the plates on the universal testing machine. Load was applied to the sample. The vertical compressive force along the sample produced tensile stress perpendicular to the vertical plane passing through the center of the sample plate. Fracture occurred along the vertical plane. Statistical analysis using *one-way ANOVA* ($p < 0.05$).

RESULTS

Table 1. Tensile strength measurement results between groups

Group	Mean (MPa)	s.d.
Negative control	2,62	0,56
1 time coating	4,51	0,24
2 times coating	5,45	0,55
3 times coating	1,41	0,62

Based on the results of tensile strength measurements between groups, it is known that the order of the largest to smallest tensile strength is the 2 times coating group, 1 time coating, negative control group and 3 times coating group.

Table 2. Normality test results (Saphiro-wilk) and homogeneity test (Levene)

Group	Sig.	Description	Homogeneity
Negative control	0,361	normal distribution	0.116
1 time coating	0,139	normal distribution	(homogeneous distribution)
2 times coating	0,785	normal distribution	
3 times coating	0,422	normal distribution	

All groups were known to be normally distributed so that they continued with the *oneway ANOVA* test with a significance level of $p < 0.05$. The results of the *oneway ANOVA* test showed a sig value. 0.000 so that there are significant differences between test groups. To determine the difference between each group, the least significant difference (LSD) post hoc test was conducted.

Table 3. Results of post hoc least significant difference (LSD) test

LSD	Negative control	1 time coating	2 times coating	3 times coating
Negative control		0,000	0,000	0,001
1 time coating			0,005	0,000
2 times coating				0,000
3 times coating				

Based on the results of the post hoc test, it is known that all test groups have significant differences in tensile strength test results.

DISCUSSION

The results showed significant differences between the negative control group and the treatment group of *Bombyx mori L.* fibroin coating on collagen membrane. The tensile strength of the treatment group was greater than the negative control group. This occurs because the polymer structure of fibroin binds to collagen. Fibroin has the same components as collagen membrane. These components are glycine polypeptides (20) making it easier for bonding to occur.

The structural strands between the fibroin polypeptide and the collagen membrane bind with hydrogen bonds. The bonding between these two materials causes an increase in the tensile strength of the collagen membrane as it is affected by fibroin which has better physicochemical properties. The superior physicochemical properties of fibroin are due to the presence of H-chains and L-chains. H-chains form discrete sheet crystals that serve as the main mechanical structure component while L-chains play a complementary mechanical role. (20). The results of this study are comparable to research stating that fibroin can increase the mechanical strength of acrylic plates in the orthopedic field. (17).

The 3 times coating group showed a

decrease in tensile strength compared to the 1 time and 2 times coating groups, even the average value of the tensile strength of this group was smaller than the negative control group. This can occur because the bond between fibroin and collagen membrane has reached a saturation point. After exceeding the culmination point, the brittle fibroin will dominate the mechanical characteristics of the collagen membrane. So that the elastic modulus of the collagen membrane decreases and then followed by a decrease in the tensile strength of the collagen membrane.

The greatest tensile strength was achieved in the 2 times coating group. This condition can occur because the treatment of 2 times coating reached the peak of maximum bonding between fibroin and collagen membrane.

CONCLUSION

It is concluded that *Bombyx mori L.* fibroin coating on collagen membrane improves the mechanical properties of tensile strength. The greatest tensile strength was achieved in the 2 times coating group.

ACKNOWLEDGEMENT

The authors would like to thank the Doctor of Dental Medicine study program, Faculty of Dentistry, Universitas Gadjah Mada for providing support through the community fund research grant program.

REFERENCES

1. Riskesdas National Report. Report_National_RKD2018_FINAL.pdf [Internet]. Agency for Health Research and Development. 2018. p. 198. Available from: http://labdata.litbang.kemkes.go.id/images/download/laporan/RKD/2018/Laporan_Nasional_RKD2018_FINAL.pdf
2. Wahjuni S, Mandanie SA. Fabrication of Combined Prosthesis with Castable Extracoronary Attachments (Laboratory Procedure). *J Vocat Heal Stud*. 2017;1(2):75.
3. Zitzmann NU, Hagmann E, Weiger R. What is the prevalence of various types of prosthetic dental restorations in Europe? *Clin Oral Implants Res*. 2007;18(SUPPL. 3):20-33.
4. Elani HW, Starr JR, Da Silva JD, Gallucci GO. Trends in Dental Implant Use in the U.S., 1999- 2016, and Projections to 2026. *J Dent Res* [Internet]. 2018;97(13):1424-30. Available from: <https://doi.org/10.1177/0022034518792567>
5. Rosenstiel SF, Land MF, Junhei Fujimoto. Contemporary Fixed Prosthodontics [Internet]. Contemporary Fixed Prosthodontics. 2016. 890 p. Available from: <http://evolve.elsevier.com/Rosenstiel/prosthodontics/>
6. Chiapasco M, Zaniboni M. Clinical outcomes of GBR procedures to correct peri-implant dehiscences and fenestrations: A systematic review. *Clin Oral Implants Res*. 2009;20(SUPPL. 4):113-23.
7. Bornstein MM, Halbritter S, Harnisch H, Weber H-P, Buser D. A retrospective analysis of patients referred for implant placement to a specialty clinic: indications, surgical procedures, and early failures. *Int J Oral Maxillofac Implants* [Internet]. 2008;23(6):1109-16. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19216281>
8. Hämmerle CHF, Jung RE, Feloutzis A. A systematic review of the survival of implants in bone sites augmented with barrier membranes (guided bone regeneration) in partially edentulous patients. *J Clin Periodontol*. 2002;29(SUPPL. 3):226-31.
9. Allan B, Ruan R, Landao-Bassonga E, Gillman N, Wang T, Gao J. Collagen membrane for guided bone regeneration in dental and orthopedic applications. *Tissue Eng - Part A*. 2021;27(5-6):372-81.
10. Elgali I, Omar O, Dahlin C, Thomsen P. Guided bone regeneration: materials and biological mechanisms revisited. *Eur J Oral Sci*. 2017;125(5):315-37.
11. Singh KP, Jayasomu RS. Bombyx mori - A review of its potential as a medicinal insect. *Pharm Biol*. 2002;40(1):28-32.
12. Nindhia TGT, Knejslik Z, Nindhia TS, Surata IW, Rumi T. Biocompatibility, Morphology, and Chemical Elements Composition of Indigenous Indonesian Wild Silkworm Cocoon for Future Application in the Field of Biomaterials. *IOP Conf Ser Mater Sci Eng*. 2020;761(1).
13. Correia C, Bhumiratana S, Yan LP, Oliveira AL, Gimble JM, Rockwood D,. Development of silk-based scaffolds for tissue engineering of bone from human adipose-derived stem cells. *Acta Biomater* [Internet]. 2012;8(7):2483-92. Available from: <http://dx.doi.org/10.1016/j.actbio.2012.03.019>
14. Kunz RI, Brancalhão RMC, Ribeiro LDFC, Natali MRM. Silkworm Sericin: Properties and Biomedical Applications. *Biomed Res Int*. 2016;2016.
15. Sunarintyas S, Siswomihardjo W, Tontowi AE. Cytotoxicity of Cricula triphenestrata cocoon extract on human fibroblasts. *Int J Biomater*. 2012;2012.
16. Miyamoto S, Koyanagi R, Nakazawa Y, Nagano A, Abiko Y, Inada M. Bombyx mori silk fibroin scaffolds for bone regeneration studied by bone differentiation experiment. *J Biosci Bioeng* [Internet]. 2013;115(5):575-8. Available from: <http://dx.doi.org/10.1016/j.jbiosc.2012.11.021>
17. Choi Y, Cho SY, Heo S, Jin HJ. Enhanced mechanical properties of silk fibroin-based composite plates for

- fractured bone healing. *Polym Fibers*. 2013;14(2):266-70.
18. Ang SL, Shaharuddin B, Chuah JA, Sudesh K. Electrospun poly(3-hydroxybutyrate-co-3-hydroxyhexanoate)/silk fibroin film is a promising scaffold for bone tissue engineering. *Int J Biol Macromol [Internet]*. 2020;145:173-88. Available from: <https://doi.org/10.1016/j.ijbiomac.2019.12.149>
19. Lim G, Lin G-H, Monje A, Chan H-L, Wang H-L. Wound Healing Complications Following Guided Bone Regeneration for Ridge Augmentation: A Systematic Review and Meta-Analysis. *Int J Oral Maxillofac Implants*. 2018;33(1):51-50.
20. Koh LD, Cheng Y, Teng CP, Khin YW, Loh XJ, Tee SY,. Structures, mechanical properties and applications of silk fibroin materials. *Prog Polym Sci [Internet]*. 2015;46:86-110. Available from: <http://dx.doi.org/10.1016/j.progpolymsci.2015.02.001>